



## Research Article

# Pharmacological effects of *Nephrolepis exaltata* L. (fern) aqueous extract on an insect-based model (*Nauphoeta cinerea*)

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## Abstract

In this work we used semi-isolated heart of the cockroach *Nauphoeta cinerea* for the investigation of the pharmacological effects of extracts (aqueous, 1:1, 1:2, 1:4 and 1:8) from *Nephrolepis exaltata* L. leaves, a popular ornamental fern considered to be safe. The use of insects in experimental studies has grown due to the easy handling, proliferation/growing assuring its rapid obtention, and absence of ethical issues. An aqueous extract 0.2 % was obtained after maceration of 1 g *N. exaltata* leaves powder with 20 mL of distilled water (1:20). Diluted extracts in water were obtained to have the following proportion 1:1, 1:2, 1:4 and 1:8. Experiments (n=4) consisted of 200 µL addition onto semi-isolated heart preparation of *N. cinerea* with concomitant heart beating counting. Aqueous, 1:1 and 1:2 extracts paralyzed completely the heart beatings of cockroaches ( $p < 0.05$  compared to saline control), but not 1:4 or 1:8, which showed only a slight decline ( $p > 0.05$  compared to saline control). A preliminary thin layer chromatography showed the presence of unidentified terpenoid in aqueous extract of *N. exaltata*. These pharmacological findings of *N. exaltata* can be exploited for future use as insecticide or as dose-dependently cholinergic agent.

## Introduction

The American fern *Nephrolepis exaltata* (L.) Schott known as 'Bostoniensis' is an ornamental plant considered to be safe [1], with few reports of contact dermatitis [2-4]. *N. exaltata* has ability to hydrolyze the acetates of phenols and aromatic-aliphatic alcohols in biotransformation reactions [5]. The plant also has been extensively studied in the metals adsorption, a process known as phytoremediation, such as arsenic [6], mercury [7], cadmium [8], and cuprum II [9]. Antimicrobial activity was reported using essential oil from aerial parts of *N. exaltata* which contain 2,4-Hexadien-1-ol (16.1%), nonanal (14.4%),  $\beta$ -Ionone (6.7%) and thymol (2.7%) [10].

The objectives of this work were: 1) to evaluate pharmacological effects of the aqueous extract of fern leaves from *Nephrolepis exaltata* on semi-isolated heart of cockroaches *Nauphoeta cinerea*, and 2) to find bioactive compounds in *N. exaltata* aqueous extract using thin layer chromatography. Our hypothesis was based on the influence of aqueous extract could have on cardiac beatings, since it is known that substances able in paralyzing the cardiac beatings could have insecticidal potential [11]. Research on cockroaches is becoming frequent by its easy maintenance, low cost, rapid reproduction and there are no restrictions on its use [12]. Besides, in recent years,

organic agriculture has become an alternative to food consumption, since the problems that insecticides can cause to human health and the environment are already known [13]. However, the extract from *Nephrolepis exaltata* tested here showed a cardiotoxic effect, with a possible effect induced by a terpene, could be used as an insecticide.

## Material and Methods

### Botanical material

The botanical material was commercially purchased from Companhia de Entrepostos e Armazéns Gerais de São Paulo (CEAGESP), in Sorocaba city, cleaned and weight (6.4 g), oven- air dried during 24 h at 40°C, and further milled to obtain 1.11 g of powder (percentage of incoming=17.15 %).

### Extracts obtention

The extract was obtained by water maceration method [14]. For each 1 g plant powder was used 20 mL distilled water, put in contact during 48 h, resulting in a 0.2% aqueous extract. Dilutions of aqueous extract in water were also performed to have 1:1, 1:2, 1:4 and 1:8 extracts.

### Biological material

Cockroaches (*Nauphoeta cinerea*) commercially purchased from Pet Shop Bicho & Cia, Sorocaba city, were put in an adequate recipient for its creation. The cockroaches had water and food (as fruits and vegetables) *ad libitum*.

### Semi-isolated heart of *Nauphoeta cinerea* preparation

The cockroaches were immobilized using entomological pinheads. The lateral margins of abdomen were cut along on each side [15] and the cuticle removed using an ophtalmic scissor to expose the visceral organs which were carefully keep out using a tweezer, in order to view the vascular system. After, the insect was put under a 30X magnifying glass (Carl Zeiss, Germany) aiming to visualize the cardiac beatings. The mean beatings/minute in the first 5 minutes were considered as zero time. After the extract addition (200 µL of each), the beatings were accounted in intervals of 5, 15, and 30 minutes [16]. Experimental groups (n=4) were randomly distributed in: saline control, pure extract, extract diluted in water 1:1, 1:2, 1:4 and 1:8. The obtained results were compared to pharmacological controls as acetylcholine chloride (110 µM, MW 181.66, Sigma-Aldrich) and adrenaline (110 µM, MW 183.20, Sigma-Aldrich), at the same 200 µL.

### Thin layer chromatography (TLC)

Aliquots of pure extract from the powder of *N. exaltata* were spotted onto 0.3mm thick silica-gel or GF254 plates (Merck®) along with appropriate standards [17, 18]. The presence of phytochemical groups such as phenolic acids and flavonoids was investigated by comparison with the following standards (Sigma): caffeic acid, chlorogenic acid, tannic acid, rutin, quercetin, atropin, β-sitosterol, coumarin, lupeol, betulin, oleanolic acid, ursolic acid, all solubilized in absolute ethanol (1mg/mL). Spots were observed with or without chromogenic agent (UV light, 360 nm). The spot of each standard was compared with spots exhibited by *N. exaltata* extract. The following systems were used as described elsewhere [19]: System I: ethyl acetate (Synth®); formic acid (Anidrol®); acetic acid (Chemco®); H<sub>2</sub>O (100:11:11:24). System II: ether (Anidrol®); toluene (Synth®) (1:1). Chromogenic agents were: A) NP/PEG: 5% (v/v) ethanolic NP (diphenylboric acid 2-aminoethyl ester, Sigma®), followed by 5% (v/v) ethanolic PEG4000 (polyethylene glycol 4000, Synth®), being visualized under UV light at 360nm. B) Sulphuric anisaldehyde (0.5 mL anisaldehyde solved in 10 mL acetic acid, 85 mL methanol and 5 mL H<sub>2</sub>SO<sub>4</sub> concentrate), heating the plaque during 10 min at 100 °C. C) Dragendorff/KOH reagent: Dragendorff (Solution "a"–2 g bismut subnitrate in 25

mL acetic acid, complete to 100 mL distilled water. Solution “b” –40 g KI, complete to 100 mL distilled water. Mixture 10 mL solution “a” and 10 mL solution “b”, add 20 mL acetic acid, complete to 100 mL distilled water); KOH 10 % diluted in ethanol.

### Statistical analysis

Results were expressed as the mean  $\pm$  standard error mean ( $\pm$ SEM). The Student's *t*-test or repeated measures ANOVA were used for statistical comparison of the data. The significance level was set at 5%.

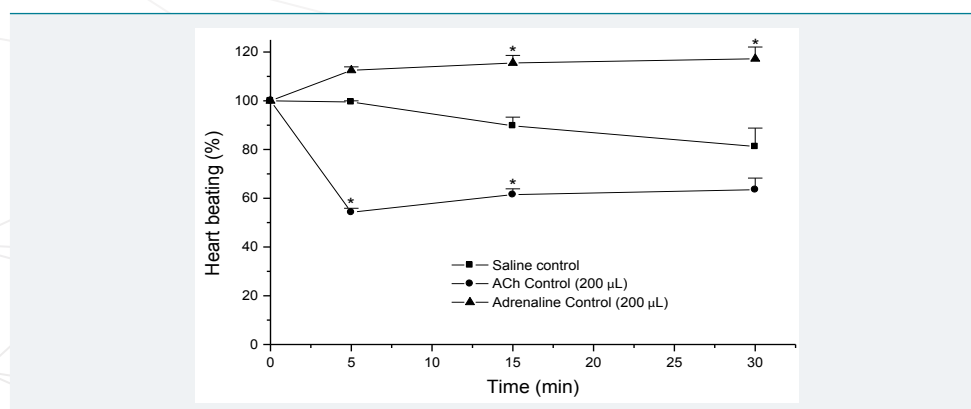
### Results and Discussion

Figure 1 expresses the percentage of heart beating (Y-axis) in function of counting in intervals of 5, 15 and 30 minutes (X-axis) of classic pharmacological drugs as adrenaline and acetylcholine in comparison to saline. Notice the effects sympathomimetic (increase of heart beatings) and parasympathomimetic (decrease of heart beatings) in semi-isolated heart of *N. cinerea*, induced by adrenaline and acetylcholine, respectively. Adrenaline is the active sympathomimetic hormone from the adrenal medulla. It stimulates both the alpha- and beta- adrenergic systems, consequently stimulates the heart [20]. Acetylcholine is a neurotransmitter found at neuromuscular junctions, autonomic ganglia, parasympathetic effector junctions, a subset of sympathetic effector junctions, and at many sites in the central nervous system [21]. At heart, acetylcholine acts causing bradycardia due to blockade of post-junctional muscarinic receptors, whereas the slight recovery seen at 15 and 30 minutes can express the vagal stimulation [22].

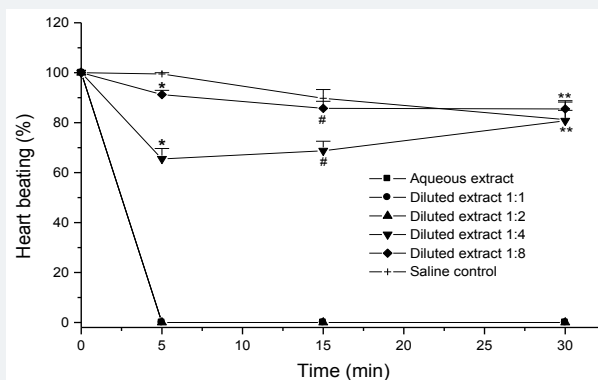
Figure 2 shows the pharmacological effects of extracts in different concentrations. The heart beating blocker effect seen with aqueous and diluted 1:1 and 1:2 extracts can be exploited for insecticidal use of the fern *N. exaltata*, which is among the most popular ornamental ferns [23].

Diluted 1:4 *N. exaltata* extract did exhibit similar profile of acetylcholine, which means that the blocker effect is a dose-dependent pharmacological event. On the other words, other than insecticidal use can be ventured to *N. exaltata*, for example, as a cholinergic agonist. However, the future pharmacodynamic of *N. exaltata* must be studied.

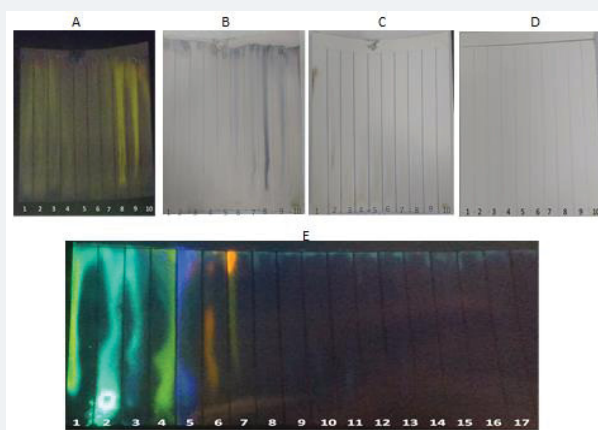
Figure 3 shows the chromatographical profile of *N. exaltata* aqueous extract face to two systems of mobile phases and three chromogenic agents, which matched to each other, resulted in the phyto group identification. Note that *N. exaltata* exhibits the same chromatographical profile of terpenes (Figure 3A, 3B) but does not as flavonoids (Figure 3E).



**Figure 1:** Semi-isolated heart preparation of *Nauphoeta cinerea*. Pharmacological effects of 200 µL addition of adrenaline and acetylcholine (both 110 µM, n=4), in comparison to SALINE CONTROL (n=4), on the exposed preparation, with sympatho- and parasympatho- mimetic effects, respectively. \* p<0.05 compared with saline control.



**Figure 2:** Semi-isolated heart preparation of *Nauphoeta cinerea*. Pharmacological effects of 200  $\mu$ L addition of *Nephrolepis exaltata* aqueous extracts (n=4, each), in comparison to saline control (n=4). Note the irreversible blockade of heart beating with aqueous and diluted 1:1 and 1:2 extracts. Diluted 1:4 and 1:8 extracts showed declines of 35 % and 9 %, respectively at first 5 minutes, with subsequent recovery at 30 min with no statistical difference in comparison to saline control. \*p<0.05, in comparison among groups and saline control. #p>0.05 (in 1:4 extract, compared to acetylcholine; 1:8 extract, compared to saline control). \*\*p>0.05 compared to saline control.



**Figure 3:** Thin layer chromatography - System I (A, B, C, and E). System II: D. Chromogenic agents: Sulphuric anisaldehyde: A (under 360 nm UV light) and B (with no UV light). Dragendorff/KOH 10%: C. NP/PEG: D and E. Standards of A, B, C, and D: 1, Atropine. 2,  $\beta$ -Sitosterol. 3, Cumarin. 4, Lupeol. 5, Betulin. 6 and 7, Oleanolic acid. 8 and 9, Ursolic acid. 10, *Nephrolepis exaltata* aqueous extract. Standards of E: 1, Caffeic acid. 2, Chlorogenic acid. 3, Tannic acid. 4, Apigenin. 5, Gallic acid. 6, Rutin. 7, Quercetin. 8, Atropine. 9,  $\beta$ -Sitosterol. 10, Cumarin. 11, Lupeol. 12, Betulin. 13 and 14, oleanolic acid. 15 and 16, Ursolic acid. 17, *Nephrolepis exaltata* aqueous extract.

Terpenes are a diverse group of more than 30,000 lipid-soluble compounds. Terpenoids are classified according to the number of isoprene units they contain. Thus, 1 unit, hemiterpene; 2 units, monoterpenes; 3 units, sesquiterpenes; 4 units, diterpenes; 5 units, sesterpenes; 6 units, triterpenes; and 8 units, tetraterpenes [24]. Terpenes exhibit a range of toxicity which confer antimicrobial properties and a wide range of effects within the insect central nervous system, but very low toxicity to mammals [25]. Phytochemical studies must be carried out in order to identify the terpene structure in aqueous extract of *N. exaltata*.

## Conclusion

Concluding, aqueous extract of fern *Nephrolepis exaltata* is cardiotoxic to cockroaches *Nauphoeta cinerea* on a dose-dependent manner. An unidentified terpene present in *N. exaltata* could contribute to this effect.

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