

Research Article

In silico analysis and characterization of fresh water fish ATPases and homology modelling

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Abstract

ATPases is known to be a crucial in many biological activities of organisms. In this study, physicochemical properties and modeling of ATPases protein of fish was analysed using *In silico* approach. ATPases a protein selected from fish species, including Gold fish (*Carassius auratus auratus*), Zebra fish (*Hypancistrus zebra*), White fishes (*Coregonus autumnalis*), Grass carp (*Ctenopharyngodon idella*) and *Anabas testudineus* (Koi) were used in this study. Physicochemical characteristics showed with molecular weight (25045.58-25148.57Da), theoretical isoelectric point (9.30-9.97), extinction coefficient(26470-34950), aliphatic index(147.31-150.35), instability index(32.84-42.67), total number of negatively charged residues and positively charged residues (5/7-6/8), and grand average of hydropathicity (1.014-1.151) were computed. All proteins were classified as transmembrane proteins. In secondary structure prediction, all proteins were composed of random coils as predominant, followed by extended strands, alpha helix and beta turn. Three dimensional structure of protein were predicted and verified as good structures. All model structures were evaluated being accepted and reliable based on structural evaluation and stereo chemical analysis.

Introduction

ATPases an enzyme that hydrolyzes ATP; *especially*: one that hydrolyzes ATP to ADP and inorganic phosphate called also *adenosine triphosphatase*. Medical Definition of ATPase an enzyme that hydrolyzes ATP; Transmembrane ATPases are membrane-bound enzyme complexes/ion transporters that use ATP hydrolysis to drive the transport of protons across a membrane. Some transmembrane ATPases also work in reverse, harnessing the energy from a proton gradient, using the flux of ions across the membrane via the ATPase proton channel to drive the synthesis of ATP. There are several different types of transmembrane ATPases, which can differ in function (ATP hydrolysis and/or synthesis), structure (e.g., F-, V- and A-ATPases, which contain rotary motors) and in the type of ions they transport. The different types include: F-ATPases (ATP syntheses, F1F0-ATPases), which are found in mitochondria, chloroplasts and bacterial plasma membranes where they are the prime producers of ATP, using the proton gradient generated by oxidative phosphorylation (mitochondria) or photosynthesis (chloroplasts). V-ATPases (V1V0-ATPases), which are primarily found in eukaryotes and they function as proton pumps that acidify intracellular compartments and, in some cases, transport protons across the plasma membrane. They are also found in bacteria. A-ATPases (A1A0-ATPases), which are found in Archaea and function like F-ATPases, though with respect to their structure and some inhibitor responses, A-ATPases are more closely related to the V-ATPases-ATPases (E1E2-ATPases), which are found in bacteria and in eukaryotic plasma membranes and organelles, and function to transport a variety of different ions across membranes'-ATPases, which are cell-surface enzymes that hydrolyse a range of NTPs, including extracellular ATP. F-ATPases (also known as ATP Synthase, F1F0-ATPase, or H (+)-transporting two-sector ATPase) are composed of two linked complexes: the

F1 ATPase complex is the catalytic core and is composed of 5 subunits (alpha, beta, gamma, delta, epsilon), while the F0 ATPase complex is the membrane-embedded proton channel that is composed of at least 3 subunits (A-C), with additional subunits in mitochondria. Both the F1 and F0 complexes are rotary motors that are coupled back-to-back. In the F1 complex, the central gamma subunit forms the rotor inside the cylinder made of the alpha (3) beta (3) subunits, while in the F0 complex, the ring-shaped C subunits forms the rotor. These ATPases can also work in reverse in bacteria, hydrolyzing ATP to create a proton gradient.

Materials and Methods

Protein sequence obtained and analysis

ATP Synthase F₀ proteins of different fish species, including Gold Fish (*Carassius auratus auratus*-Accession No. -AEH99465.1), *Hypancistrus zebra* (Accession No. APF31803.1), *Coregonus autumnalis* (Accession No. -ABO14995.1), *Ctenopharyngodon idella* (Accession No. -ALS20290.1), *Anabas testudineus* (Accession No- AQT00813.1) were retrieved from the NCBI Protein database (<http://www.ncbi.nlm.nih.gov>) under the FASTA format for analysis. The above obtained sequence was further used for complete protein analysis (structure and functional annotation) and model building using comparative modelling approach. Using expasy's protparam server (<http://expasy.org/cgi-bin/protparam>) complete primary structure analysis has been performed. SOPMA was used for secondary structure prediction of protein sequence.

Sequence alignment

Multiple sequence alignment between Mitochondrion ATPase sequences from different fish species was performed using the clustalW2 server (<http://www.ebi.ac.uk/tools/msa/clustalW2/>). Neighbour- Joining phylogenetic analysis of protein sequences was also generated using Clustal omega [1-3]. Clustal W2 is a server for multiple sequence alignment which is also used for phylogenetic tree analysis. Phylip and mega also available server for phylogenetic tree analysis server.

Physiochemical characterization

Physiochemical properties of the proteins such as molecular weight (Mol. wt.), amino acid composition, theoretical isoelectric point (pI), total number of positive (Arg+Lys) and negative (Asp+Glu) residues (+R/-R), extinction co-efficient (EC), instability index (AI), and grand average of hydropathicity (GRAVY) of investigated proteins was analysed by searching on the Expasy's protparam server (<http://web.expasy.org/protparam/>) For the domain structures the simple Molecular Architecture Research tool (SMART) program (<http://smart.embl-heidelberg.de/>) was used, for primary structure analysis as expasy's protparam server.

Functional analysis

The server SOSUI (Hirokawa et al.) was performed to identify the types of protein. The CYS_REC (<http://linux1.softberry.com>) was used to predict the Presence or absence of disulphide bonds and their bonding pattern, which are crucial in defining the functional linkage and the stability of a protein. So, CYS_REC used to determine presence or absence of cystein bond.

Protein structure prediction

Secondary structure of proteins was predicted using SOPMA server (http://npsapabi.ibcp.fr/cgi-bin/npsa_automat.pl?page=/NPSA/npsa_sopma.html), with the default parameters (window width: 17; similarity threshold: 8; number of states: [4,5]. Homology modeling was constructed using Swiss model server (<http://swissmodel.expasy.org/>) [6,7]. Swiss model is a server which is used for 3D structure prediction and also template selection, template is select based on maximum similarity or identity

with sequence. Quality and accuracy with validation of the predicted models were analysed performing RAMPAGE for Ramachandran plot analysis [10,11]. The best selected models were based on the total number of residues in the most favoured regions, additional allowed region, generously allowed region and disallowed region as well as an overall G-factor have over 90% in most favoured region and cut off value (>-0.5) of overall G-factor [11,12]. Raptor X also available server for 3d structure prediction and PSIPRED and GOR IV also available free server for secondary structure analysis server. For protein structure prediction also available server are phyre 2, HHpred, modeller, CPH models, Iomates, Modbase and Robette etc.

Results and Discussion

Physico-chemical characterisation

Amino acid composition in ATPase computed using ExPASy's ProtParam server. The physicochemical characterisations of proteins were obtained analysing ExPASy's ProtParam tools (Table 1). The value of isoelectric point (pI) of proteins were ranged from 9.30-9.97 (more than 7), implying the basic character of these proteins. The pI values function in protein purification by isoelectric point focusing on a polyacrylamide gel. Total number of positively (Asp+Glu) and negatively (Arg+Lys) charged residues (+R/-R) was ranged from 5 to 6 and 7 to 8, respectively. The extinction co-efficient (EC) of proteins measured at 280 nm was in a range of 31970 to 34950 M⁻¹.cm⁻¹ (assuming all pairs of cysteine residues from cysteins). The high value of ECs in this study implied a high concentration of cysteine along the protein sequences, functioning in quantitate the protein concentration in a volume of solution. The Instability index (II) value evaluates the stability of proteins in a test tube; it was recommended that a protein is stable when its II value is smaller than 40 and as unstable when such value is above 40. This study results showed that the II value proteins was in a range of proteins was in a range of 32.84-42.67 showing the protein of zebra fish is (II>40) and the rest is stable (II<40) [13-22]. The aliphatic index (AI) is a parameter for estimating thermal stability of a protein directly associating with the mole fraction of aliphatic side chains (Alanine, isoleucine, leucine and valine) in the protein. In this study a high Aliphatic index values of proteins (147.31-150.35) imply high thermo stability of these proteins. Low grand average hydropathicity (GRAVY) regarded as a measure for the stability of globular protein at high temperature. The amino acid composition in ATP Synthase-F₀ computed using ExPASy's ProtParam was showed in table 1.

All proteins were classified as transmembrane proteins through SOSUI program. The transmembrane regions predicted from protein sequences were shown in Table 2. These amino acid sequences of Membrane Protein have 6 transmembrane helices, except *Coregonus autumnalis* which have 5 transmembrane regions.

Sequence alignment

A multiple amino acid sequence alignment of proteins was performed [1] (Figure 1A,B). The result indicated a high amino acid sequence similarity between the ATPases of five studied fish species; it was observed that between *Coregonus* and *Hypancistrus* was greater similarity. A neighbour-joining phylogenetic tree was constructed using Clustal Omega.

Table 1: Parameters computed using ExPASy's ProtParam tool.

Species	No. of aa	Mol. wt.	pI	+R/-R	EC	II	AI	GRAVY
Zebra fish	227	25148.57	9.30	6/8	31970	42.67	149.56	1.014
Grass Carp	227	25055.57	9.51	5/7	31970	32.84	147.31	1.151
Autumnalis	227	25055.57	9.51	5/7	31970	32.84	147.31	1.151
Gold fish(auratus)	227	25047.49	9.30	5/7	34950	33.16	150.35	1.135
Anabas testudineus	227	25045.58	9.97	5/8	26470	36.10	147.44	1.080

Table 2: Types of protein and transmembrane region identified by using SOSUI.

Species	Types of protein	ength	Transmembrane region	N-C terminal
Grass Carp	PRIMARY	23	PSFLGIPLIAIAIALPWVLFPTP	60-82
	SECONDARY	23	WALLLASLMVFLITINMLGLLPY	117-139
	PRIMARY	20	SLNMGFAVPLWLATVIIGMR	148-167
	PRIMARY	23	PLIPVLIIETISLIRPLALGV	185-207
	PRIMARY	23	TAGHLLIQLIATAVAVFVLLPMMPT	214-236
Zebra fish	PRIMARY	23	FLTLLEAVAVAMIQAYVFVLLLS	246-268
	PRIMARY	22	PTFLGIPLIAIALTLPWILIPS	55-76
	SECONDARY	23	WALITSLMIFILSLNMLGLLPY	112-134
	PRIMARY	20	LSLNMGFAVPLWLATVIIGL	142-161
	PRIMARY	23	LIPVLIIETISLIRPLALGVR	181-203
Anabas testudineus	SECONDARY	23	HLLIQLISTATFILLPMMTTVAL	212-234
	PRIMARY	23	ILLTLEAVAVAMIQAYVFVLLLS	241-263
	PRIMARY	23	PIFLGVPLIALALALPWILFPTP	55-77
	SECONDARY	23	WALLFTSLMLFLMTLNMLGLLPY	112-134
	SECONDARY	21	LSLNMAFAVPLWLATVIIGMR	142-162
Carassius auratus	PRIMARY	23	PLIPVLIIETISLIRPLALGV	180-202
	PRIMARY	23	IQLIATAAFVLLPLMPAVAILTA	215-237
	PRIMARY	22	IQLIATAAFVLLPLMPAVAILTA	248-269
	SECONDARY	23	ASPSYLGIPLIAIAIALPWVLYP	57-79
	SECONDARY	23	WALLLASLMIFLITINMLGLLPY	116-138
Coregonus autumnalis	PRIMARY	20	SLNMGFAVPLWLATVIIGMR	147-166
	PRIMARY	23	PLIPVLIIETISLIRPLALGV	184-206
	PRIMARY	23	TAGHLLIQLIATAVAVFVLLPMMPT	213-235
	PRIMARY	23	FLTLLEAVAVAMIQAYVFVLLLS	245-267
	SECONDARY	23	ISFMSPTYLGIPLIAVALTLPWII	47-69
Coregonus autumnalis	PRIMARY	23	MLTSLMFLITLNMLGLLPYTFT	112-134
	SECONDARY	21	QLSLNMGAVPMWLATVIIGM	138-158
	PRIMARY	23	PLIPVLIIETISLIRPLALGV	177-199
	PRIMARY	23	IATAAFVLLPMMPTVAILTALVL	215-237

In grass carp Total length: 275 A. A. Average of hydrophobicity: 0.844728, in zebra fish Total length: 270 A. A., Average of hydrophobicity: 0.754445, Anabas testudineus, Total length: 270 A. A., Average of hydrophobicity: 0.798519, Carassius auratus, Total length: 274 A. A., Average of hydrophobicity: 0.861314, Coregonus autumnalis, Total length: 246 A. A. Average of hydrophobicity: 0.841057.

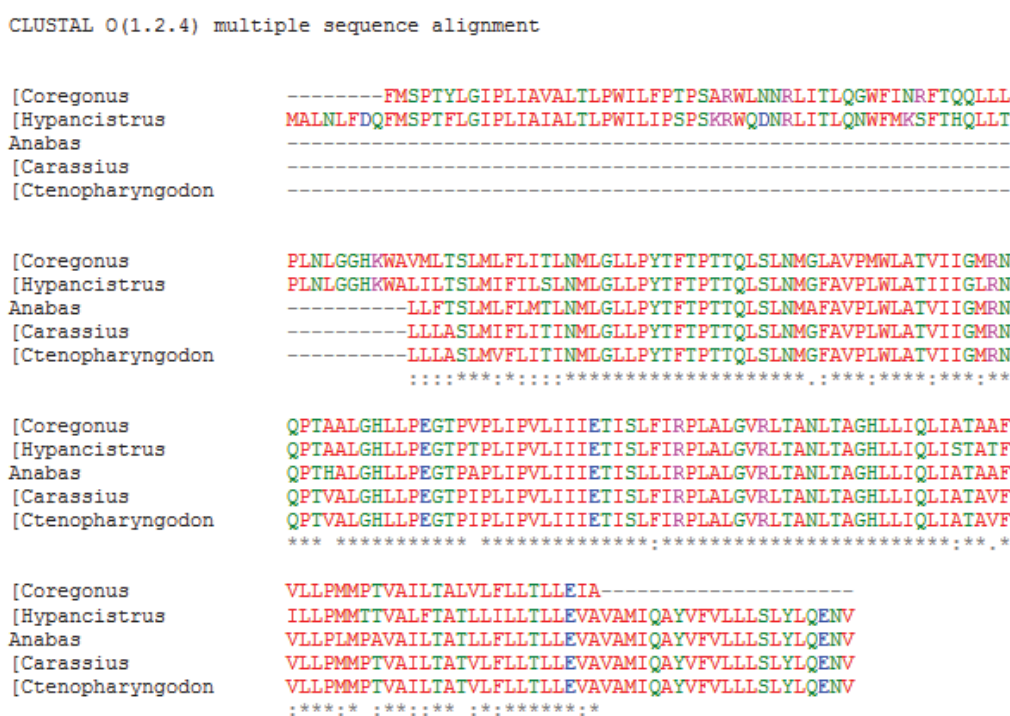


Figure 1A: Sequence alignment and phylogenetic analysis between fish ATPases sequences. (A) Multiple sequence alignment. The asterisk marks (*), colon (:), dot (.) and dash (-) indicated identical amino acids, conserved substitutions, semi conservative and deletions, respectively.

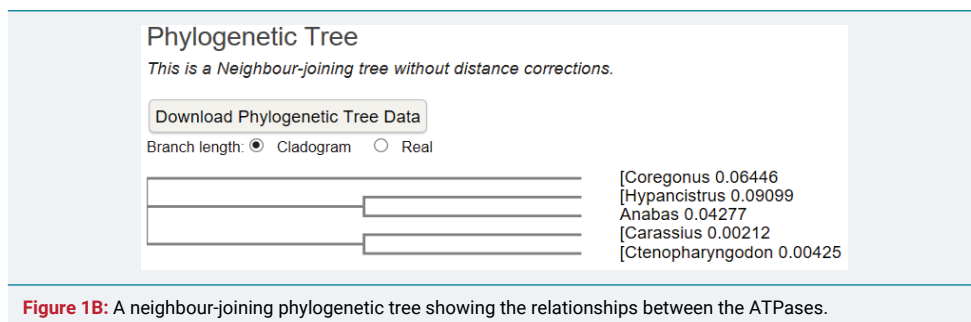


Figure 1B: A neighbour-joining phylogenetic tree showing the relationships between the ATPases.

Functional analysis result

Disulphide bonds are significant in the protein folding and stability, which are generated between the thiol groups of cysteine residues by oxidative folding process. In this study, the cysteine residues in the proteins were determined using CYS_REC server. The results revealed that any of these proteins not contain cysteine residues and most probable patterns of pairs of cysteine were not found (Table 3), suggesting that no one proteins contain disulphide bonds [4].

There is no CYS_REC found in five Fish species. Cysteine residues and disulphide bonds which are important in determining the thermo stability of proteins. The results indicated the probable absence of disulphide bonds in these proteins.

Protein structure prediction and validation

The secondary structure of ATPases protein from fish species was predicted using SOPMA (Table 4). The results showed that except for ATPases from grass carp all contain alpha helix as a predominant component among the secondary structure elements, followed by random coil, extended strand and beta turn.

The three dimensional structures of fish ATPases protein were modelled based on the sequence and structural similarity to different available protein structure templates from the pdb (Table 4). The final structure of the models represented with the Swiss pdb viewer was shown in figure 2. Validation and predicted models performing Rampage for Ramachandran plots were represented in table 5 [4].

The stereo chemical quality and accuracy of proposed models were examined performing PROCHECK analysis shown in table 4. The analysis results revealed that the predicted models for ATPase of Grass Carp, zebra fish, Anabas testudineus, Carassius auratus, and Coregonus autumnalis have over (88%) of residues in the most favoured region, indicating that these homology models were good quality and additional allowed regions combined, implying acceptable. Results showed that over 88% (88.8%, 88.4%, 88.4%, 88.5%, and 88.6% respectively) of residues found in the most favoured regions. More over 7% (7.7%, 7.6%, 7.6%, 7.7%, 7.7%) of residues in the generously or additional allowed regions, and (3.5,4.0,4.0,3.8,3.8)% residues in the disallowed regions of the proteins. All protein models contained the lower than 8% fell of residues in the generously allowed regions, indicating that may be near to be good quality models. Q-mean score value range (-5.68-5.34), the result implied that models were accepted.

Conclusion

In this study, five ATPase proteins of freshwater fish species were selected to characterise using computational tools. Physicochemical and functional characterisations of the proteins were profoundly investigated. All proteins were classified as transmembrane protein, an approximately number of alpha helix and random coils were computed to be dominating, followed by extended strands in the secondary structure of all proteins. The three dimensional models of proteins

Table 3: Disulphide bond patterns of pairs predicted using CYC_REC tool.

Species	CYS_REC
grass carp	Nil
zebra fish	Nil
Anabas testudineus	Nil
Carassius auratus	Nil
Coregonus autumnalis	Nil

Table 4: Secondary structure elements of ATPases of fish species using SOPMA.

Element	Grass Carp	zebra fish	Anabas testudineus	Carassius auratus	Coregonus autumnalis
Alpha Helix	113(49.78%)	120(52.86%)	125(55.95%)	111(48.90%)	92(46.46%)
3 ₁₀ Helix	0(0.00%)	0(0.00%)	0(0.00%)	0(0.00%)	0(0.00%)
Pi helix	0(0.00%)	0(0.00%)	0(0.00%)	0(0.00%)	0(0.00%)
Beta Bridge	0(0.00%)	0(0.00%)	0(0.00%)	0(0.00%)	0(0.00%)
Extended Strand	36(15.86%)	36(15.86%)	29(12.78%)	39(17.18%)	37(18.69%)
Beta Turn	7(3.08%)	7(3.08%)	11(4.85%)	11(4.85%)	6(3.03%)
Bend region	0(0.00%)	0(0.00%)	0(0.00%)	0(0.00%)	0(0.00%)
Random coil	71(31.28%)	64(28.19%)	60(26.43%)	66(29.07%)	63(31.82%)
Ambiguous states	0(0.00%)	0(0.00%)	0(0.00%)	0(0.00%)	0(0.00%)
Other states	0(0.00%)	0(0.00%)	0(0.00%)	0(0.00%)	0(0.00%)

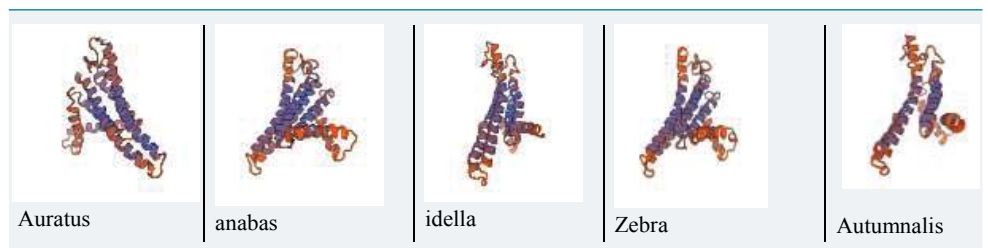


Figure 2: Structure of the models represented with the Swiss pdb viewer of different fish protein.

Table 5: Homology modeling of three –dimensional (3-D) structures of ATPase protein of fish species using SWISS MODEL and Rampage for Ramachdran plot analysis.

Index	Grass carp	zebra fish	Anabas testudineus	Carassius auratus	Coregonus autumnalis
Template	5arh.1.V	5ara.1.V	5ara.1.V	5are.1.V	5ara.1.V
Resolution (Å)	7.2	7.2	6.7	7.4	6.7
Sequence identity (%)	62.42	53.49	60.51	55.38	52.31
Q mean	-3.71	-5.68	-3.82	-4.37	5.34
Rampage					
Residues in most favoured region	4064(88.8%)	4047(88.4%)	4047(88.4%)	4050 (88.5%)	4050(88.5%)
Residues in generously allowed region	351(7.7%)	347(7.6%)	347(7.6%)	351 (7.7%)	351(7.7%)
Residues in out lier region	161(3.5%)	182(4.0%)	182(4.0%)	175 (3.8%)	175(3.8%)

was predicted and validated by the accuracy of ramachdran plot analysis; the results suggested that all proposed models are reliable and valid. This study provide information on the physiochemical characteristics, structural properties and molecular functions of fish ATPase, which are useful for further studies on specific functions.

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