Molecular analysis of immunoglobulins related to salmonella typhi in pediatric patients

Farah Qudsia¹, Muhammad Rehan² and Samreen Riaz³*

¹Department of Microbiology and Molecular Genetics, University of the Punjab, Lahore, Pakistan
²Research Officer, IoT Research and Innovation Lab, Al-Khwarizmi Institute of Computer Sciences, University of Engineering and Technology, Lahore, Pakistan
³Assistant Professor, Department of Microbiology and Molecular Genetics, University of the Punjab, Lahore, Pakistan

Introduction

Salmonella enterica infections are a unifying common cause of bloodstream infection in developing countries. In these areas, it is related to a high case fatality quantitative relation [1]. Typhoid, caused by the Gram-negative bacterium Salmonella enterica serovar Typhi, could be a major explanation for community-acquired bacteremia and death worldwide [2]. Various clinical presentations are associated with it which also overlap many other acute febrile illnesses [3]. Noticeable cases of enteric fever caused by Salmonella enterica serovars Typhi and Paratyphi are routinely reported which have high impact in toddlers and youngsters [4]. It is a common childhood problem in both urban and rural regions in tropical countries. Especially Asia, Africa, Eastern Europe, and Latin America are at high risk [3]. Asia is mostly affected by it contributing 93% of the global episodes. The major causative factors are poor hygiene, poor sanitation and sewerage system and over population [5]. The predominant mode of transmission was believed to be the direct fecal-oral route but recent proof suggests that indirect environmental transmission may additionally contribute in spreading the disease [6]. It is still an increasing problem in under developed countries like Pakistan [7]. Salmonella enterica has initially acquired antimicrobial resistance against the traditional first-line drugs like chloramphenicol, ampicillin, and trimethoprim-sulfamethoxazole [1]. The major cause of the emergence of antibiotic resistance is late treatment of typhoid fever due to incorrect diagnostic assay results and their interpretation. The arrival of a speedy check for identification of typhoid has returned as a blessing for clinicians and patients [8]. Typhoid fever cannot be diagnosed by the physician on the basis of clinical features as they are very similar in common infections like malaria, dengue, typhoid, leptospirosis and scrub typhus [9]. An early and correct identification is important for a prompt and

Abstract

Typhoid fever is a systemic infection caused by Salmonella enterica serotype typhi. It is of major concern in tropical regions of the world. Highest episodes of typhoid fever occur in Asia i.e.93%. Early diagnosis of the disease is mandatory to lower the mortality rate associated with it as well as to prevent the emergence of antimicrobial drug resistance by Salmonella typhi. Research work was conducted in Immunology Department of the Children’s Hospital, Lahore for the period of one year including a total of 60 patients suspected of having typhoid fever. Serum samples of these patients were tested for Typhidot IgG and IgM antibodies as well as for the antibodies against TO and TH antigens using Widal test. Of the total 60 patients, 10 (16.7%) were positive for both Typhidot IgG and IgM, 16 (26.7%) were positive for Typhidot IgM, 3 (5%) were Positive for Typhidot IgG and 31 (51.66%) were negative for both Typhidot IgG and IgM. Reading the results of Widal test, 8 (13.33%) were positive for Widal TO and TH antigens, 3 (5%) were positive for Widal TO antigen, 19 (31.7%) were positive for Widal TH antigen and 30 (50%) were negative for Widal TO and TH antigens. IgM is positive at the early stage of acute typhoid fever, IgM along with IgG positive means the middle stage of acute illness. The detection of only IgG cannot discriminate between acute and convalescent phases as it can stay in the serum for at least 2 years or more. The Typhidot test is much helpful for the rapid diagnosis of typhoid fever as compared to Widal test which is still being used in some set ups in poor countries, although has become mostly obsolete. By testing the rise of IgM and IgG antibodies against Salmonella typhi, we can detect the infection at early and late stages, respectively.

More Information

*Address for Correspondence: Samreen Riaz,
Assistant Professor, Department of Microbiology and Molecular Genetics, University of the Punjab, Lahore, Pakistan.
Email: samreen.mmg@pu.edu.pk

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A simple, reliable, affordable, and speedy diagnostic assay has been a long-felt requisite of the clinicians [11]. Form always for its identification are blood culture and Widal test. The gold standard for Typhoid fever is blood culture but is time consuming [12]. Also, the results of the blood culture are affected by prior administration of antibiotics. Widal test gives positive results after increasing titers and this is often not helpful for early medical care [13].

Typhidot is a speedy serological test that is now commercially accessible and reliable in identification of typhoid fever even with restricted resources [14]. It is a dot enzyme immunoassay (EIA) that identifies the immunoglobulins IgM and IgG against the specific 50KD outer membrane protein (OMP) of the Salmonella typhi. Typhidot has been used as the sole tool to diagnose typhoid fever by general practitioners and consultants [7]. It is reasonable and a reliable diagnostic test for identification of typhoid in resource-limited setting among patients with a very high index of suspicion of infectious disease when blood culture is not accessible or possible [8].

Aims and objectives

Keeping in view the above literature, aims and objectives of research work were to determine the Typhidot IgG and IgM antibodies as well as the titer antibodies against TO and TH antigens using Widal test in patients suspected of having typhoid fever.

Inclusion criteria

Patients of either sex with age range of 1 year to 14 years.

Exclusion criteria

Patients with fever with some definite focus of infection.

Materials and methods

During a period of one year, all patients having a clinical suspicion of typhoid fever, reporting to the Department of Immunology, Children’s Hospital Lahore, were included in this study. The patients were children between ages of 1 to 14 years and of both sexes with clinical features suggestive of typhoid fever.

Data collection and clinical history

For data collection and clinical history, a proforma was developed. The parameters taken into consideration for recording the clinical history were fever, weakness, anorexia, abdominal pain, nausea, vomiting, cough, chest discomfort, sore throat and coated tongue.

Sample collection

5 c.c. of blood was collected aseptically from the anti-cubical vein of each patient. The blood samples were centrifuged and the serum separated was divided into two parts.

Tests performed

Typhidot and Widal tests were performed on the serum sample for the serological diagnosis of typhoid fever. The kit used for Typhidot test was “Malaysian Biodiagnostic Research SDN BHD, Singapore, Malaysia”. Whereas Widal test was performed using “Sanofi qualitative agglutination test kits (Bio-Rad)”. The tests were performed according to manufacturer’s instruction.

Protocol of typhidot test

To perform Typhidot test the reaction tray was divided into two columns. One was marked as “M” and one column as “G”. Predotted antigen strips were removed using the forceps and placed into the appropriate reaction wells. 250 μl of the sample diluent was added into the appropriate “M” and “G” reaction wells. Tray was shaked gently to allow strips to be thoroughly wet. 2.5 μl of either control or test serum was added to the appropriate “M” or “G” reaction wells to achieve a final serum dilution of 1:100. The solution was aspirated gently to mix. The tray having solutions was incubated on a rocker platform for 20 minutes. The antibody solution was aspirated into a discard jar containing disinfectant. 250 μl of prepared washing buffer was added into each well and washed 3X for five minutes. 250μl of prediluted Anti-Human IgM was added to the “M” well and 250 μl of prediluted Anti-Human IgG into the “G” well using sterile micropipette tips. The tray was covered with a cover for 15 minutes at room temperature on a rocker platform. The 2nd antibody solution was aspirated and washed 3X for 5 minutes. 250 μl of the color development solution was added into each well. The tray was covered and incubated on the rocker platform. Fifteen minutes were allowed for colour development. The reaction was stopped by aspirating the solution and briefly rinsing the strip in distilled water (3X). Strips were placed on the filter paper to dry and then results were interpreted as:

Negative result: Antigen coated strip was as blank as it was before starting the reaction.

Positive result: Antigen coated strip was showing two blue dots on it. A positive IgM was inferred as acute typhoidal illness, while IgM and IgG positive were meant for acute typhoidal illness in middle stage of infection and IgG positive was inferred as chronic carrier or previous infection or reinfection.

Protocol for widal test

To perform Widal test, the test reagents and sample were brought to the room temperature. Antigen vials were re-suspended gently. Clear glass slides were taken. For each specimen to be tested, drops of 80, 40, 10 and 5 μl of the patient’s serum were placed into separate circles of slides (for making the dilution of 1/20, 1/40, 1/80, 1/160 and 1/320, respectively). A 40 μl drop of the appropriate antigen
Results

The study included a total of 60 patients both males and females, suspected of having typhoid fever, who reported to the Children's Hospital Lahore and were referred to Immunology Department for serological diagnosis of typhoid fever.

In the study population (n = 60) of pediatric patients suspected of having typhoid fever 38 (63%) were males while 22 (37%) were females with a male: female ratio of 1.7:1 (Figure 1a).

Of the total 60 patients, 10 (16.66%) were positive for both Typhidot IgG and IgM, 16 (26.66%) were positive for Typhidot IgM, 3 (5%) were positive for Typhidot IgG and 31 (51.66%) were negative for both Typhidot IgG and IgM (Figure 1b).

A total of 10 patients with Typhidot positive results were identified, in these cases, equal number of males and females was affected. In 16 Typhidot IgM positive patients, 11 (68.75%) were males and 5 (31.25%) were females. In the Typhidot IgG positive patients all were females (100%). Among rest of the 31 Typhidot IgG and IgM negative patients 23 (74.19%) were males and 8 (25.80%) were females. Out of total patient population (n = 60), based on duration of fever, 11 (18.33%) patients were having fever for < 7 days, 23 (38.33%) were having fever for 7-14 days and 26 (43.33%) were having fever for > 14 days (Figure 2a).

Amongst patients having fever for < 7 days 2 (3.33%) were positive for both Typhidot IgG and IgM, no patient (0%) was positive for either Typhidot IgG or Typhidot IgM alone while 9 (15%) were negative for both Typhidot IgG and IgM. Amongst patients having fever for 7-14 days, 4 (6.66%) were positive for both Typhidot IgG and IgM, 7 (11.66%) were positive for Typhidot IgM, no patient (0%) was positive for Typhidot IgG, while 12 (20%) were negative for both Typhidot IgG and IgM. Amongst patients having fever for > 14 days, 4 (6.66%) were positive for both Typhidot IgG and

Figure 1: a: Gender distribution of paediatric patients, b: Results of Typhidot IgG and IgM for patients suspected of having typhoid fever (n = 60).

Figure 2: a: Duration of fever in patients suspected of having typhoid fever (n = 60), b: Results of Typhidot IgG and IgM according to duration of fever in patients suspected of having typhoid fever.
IgM, 9 (15%) were positive for Typhidot IgM, 3 (5%) were positive for Typhidot IgG and 10 (16.66%) were negative for both Typhidot IgG and IgM (Figure 2b).

In the present study, serological diagnosis of typhoid fever was also made using Widal TO and TH antigens. The results show that, of the total patients (n = 60), 8 (13.33%) were positive for Widal TO and TH antigens, 3 (5%) were positive for Widal TO antigen, 19 (31.66%) were positive for Widal TH antigen and 30 (50%) were negative for Widal TO and TH antigens (Figure 3a).

Out of 8 patients positive for Widal TO and TH antigens, equal number of males and females was affected. In 3 patients positive for Widal TO antigen all (100%) were males and amongst 19 patients positive for Widal TH antigen 10 (52.63%) were males and 9 (47.36%) were females. Among rest of the 30 Widal TO and TH antigen negative patients 23 (76.66%) were males and 7 (23.33%) were females. Amongst patients having fever for < 7 days, 1 (1.66%) was positive for Widal TO and TH antigens, no patient (0%) was positive for Widal TO antigen, 2 (3.33%) were positive for Widal TH antigen and 8 (13.33%) were negative for Widal TO and TH antigens. In patients having fever from 7-14 days, 1 (1.66%) was positive for Widal TO and TH antigens, 2 (3.33%) were positive for Widal TO antigen, 5 (8.33%) were positive for Widal TH antigen and 15 (25%) were negative for Widal TO and TH antigens. In patients having fever for > 14 days, 6 (10%) were positive for Widal TO and TH antigen, 1 (1.66%) was positive for Widal TO antigen, 12 (20%) were positive for Widal TH antigen and 7 (11.66%) were negative for Widal TO and TH antigens (Figure 3b).

In the present study, serological diagnosis was made according to set of symptoms and findings at physical examination. The common clinical presentation in patients suspected of having typhoid fever (n = 60) were fever 60 (100%), abdominal pain 45 (75%), nausea 38 (63.33%), vomiting 40 (66.66%), diarrhea 35 (58.33%), constipation 15 (25%), cough 40 (66.66%), chest discomfort 34 (56.66%), sore throat 37 (61.66%), coated tongue 42 (70%), anorexia 54 (90%) and weakness 58 (96.66%) (Table 1).

### Discussion

Typhoid is considered as an illness of past by people who are residents of developed countries. It is a systemic, mortal and contagious disease caused by *Salmonella typhi*. This bacterium specifically affects humans in a highly-adapted manner. It is found more habitually in underdeveloped countries where overload of population and improper sanitation are common [15].

Molecular Immunology has made a headway to the discovery of sensitive and specific markers of *Salmonella typhi*. They are used to make such kits which are economical and factual for speedy serological interpretation of typhoid

<table>
<thead>
<tr>
<th>Clinical presentations</th>
<th>Patients with clinical presentations</th>
<th>Patients without clinical presentations</th>
<th>Total number of patients (n = 60)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fever</td>
<td>60 (100%)</td>
<td>0 (0%)</td>
<td>60 (100%)</td>
</tr>
<tr>
<td>Abdominal pain</td>
<td>45 (75%)</td>
<td>15 (25%)</td>
<td>60 (100%)</td>
</tr>
<tr>
<td>Nausea</td>
<td>38 (63.33%)</td>
<td>22 (36.66%)</td>
<td>60 (100%)</td>
</tr>
<tr>
<td>Vomiting</td>
<td>40 (66.66%)</td>
<td>20 (33.33%)</td>
<td>60 (100%)</td>
</tr>
<tr>
<td>Diarhea</td>
<td>35 (58.33%)</td>
<td>25 (41.66%)</td>
<td>60 (100%)</td>
</tr>
<tr>
<td>Constipation</td>
<td>15 (25%)</td>
<td>45 (75%)</td>
<td>60 (100%)</td>
</tr>
<tr>
<td>Cough</td>
<td>40 (66.66%)</td>
<td>20 (33.33%)</td>
<td>60 (100%)</td>
</tr>
<tr>
<td>Chest discomfort</td>
<td>34 (56.66%)</td>
<td>26 (43.33%)</td>
<td>60 (100%)</td>
</tr>
<tr>
<td>Sore throat</td>
<td>37 (61.66%)</td>
<td>23 (38.33%)</td>
<td>60 (100%)</td>
</tr>
<tr>
<td>Coated tongue</td>
<td>42 (70%)</td>
<td>18 (30%)</td>
<td>60 (100%)</td>
</tr>
<tr>
<td>Anorexia</td>
<td>54 (90%)</td>
<td>6 (10%)</td>
<td>60 (100%)</td>
</tr>
<tr>
<td>Weakness</td>
<td>58 (96.66%)</td>
<td>2 (3.33%)</td>
<td>60 (100%)</td>
</tr>
</tbody>
</table>
fever [16]. The serotype Typhi has a specific outer membrane protein (OMP) which is of 50kDa. It was detected by Ismail, et al. who also checked out this antigen by dot-enzyme immunoassay (EIA) technique and noticed that this antigen can react with immunoglobulins M and G produced due to Salmonella typhi in the serum of the victims of acute typhoid fever [17]. The OMP is evident from the other antigens of the bacterium which are somatic (O), flagellar (H) or capsular (VI). Interpreting from the current writings Typhidot is the most suitable test for confirmation of typhoid fever. Typhidot IgM positive means early phase of illness while both IgM and IgG are positive in case of middle phase of infection. IgG antibodies are not indicative of acute infection as they persist in serum of the patient for about 2 years of infection [12]. The results of Typhidot may vary topographically because of the distinctions in the genomic structure and flexibility of the outer membrane protein of Salmonella typhi [18]. While comparing the results of Typhidot (as given in figure 1b), another study including 65 patients showed positive results for both IgG and IgM in 39 samples, IgM was positive in 24 samples and IgG in 2 samples. Hence reporting that Typhidot is 100% sensitive and 80% specific as compared to blood culture [19].

The present study showed a notable pattern when the results were arranged according to the duration of fever (Figures 2a,b and 3b). It is comparable to the findings by Jaffery, et al. which reported that amongst the patients having fever for < 7 days (n = 22), 10 (55.56%) were positive for Widal and 12 (66.67%) were positive for Typhidot. In cases with 7-14 days of illness (n = 83), 38 were positive for Widal and 45 (80.36%) were positive for Typhidot. Whereas in patients having illness of > 14 days duration (n = 44), 24 (82.76%) were positive for Widal and 20 (68.97%) were positive for Typhidot. The Typhidot showed an upper value of sensitivity and specificity to that of Widal test [16]. In another evaluation, the results of Typhidot showed high sensitivity during the first seven days of febrile illness. Seemingly, the reason for this is that while performing Typhidot, we are mainly concerned with the results of IgM which is the first antibody to be released after any infection. The level of IgG will be detectable later. In the Widal test, O agglutinins are observed at around day 8, while, H agglutinins are observed at about days 10-12 [20].

Underdeveloped countries have been using Widal test for more than hundred years but its advantage as a diagnostic tool is restricted because of less sensitivity, specificity and positive predictive value [21]. Sensitivity is reduced because Widal test may be positive after a long latent period. Specificity is reduced if the infection is repeated or patient has administered TAB vaccine or the ability to react with the other gran negative infections. When the blood culture test is combined with any serological test then it usually gives better diagnostic results specifically in those cases which are partially treated. This behavior could be tested by combining Typhidot with blood culture of those children who have already received antibiotics [22]. Routinely recommended Widal test usually does not give positive results in the first week of infection as it is dependent on increased titres which may take about 48 hours. The gold standard for typhoid is culture which is 100% specific and detect the antibiotic sensitivity. Its disadvantage is again a delay in the interpretation of results i.e. 72 hours. The results of culture are badly affected if there is prior administration of antibiotics. To overcome these drawbacks, serological tests like Typhidot and Tubex have been discovered in the recent past. These tests are helpful in the early diagnosis of typhoid fever by identifying antibodies against Salmonella typhi. Typhoid fever can be diagnosed within 20 minutes by using Typhidot test. The usefulness of Typhidot is quite profound in the diagnosis of typhoid fever. It is easy to perform, and results are interpreted quickly [12]. There is no need of an ELISA reader to evaluate the results. They are easily reported by seeing the dots on the test strips. It can easily be performed without much operator training. The hindrances in advising this test in developing countries are the cost of the test which is more than Widal test and the test reagents must be placed in cold-storage [22].

**Conclusion**

For the diagnosis of typhoid fever, Typhidot is much better than Widal test as it is more sensitive, easy to perform, detects the cases at early stage and results are available on the same day. And in Typhidot test, IgM is more prevalent than IgG meaning that more are the cases of acute infection as compared to chronic infection.

**Ethical approval**

Ethical approval was taken from Children’s hospital. An informed consent was taken from all the subjects involved.

**Acknowledgement**

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